

Photoinduced DNA cleavage by formation of ROS from oxygen with a neurotransmitter and aromatic amino acids†

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UV-B photoirradiation of a neurotransmitter (serotonin) and aromatic amino acids (tryptophan and tyrosine) with oxygen results in DNA cleavage by generation of reactive oxygen species (ROS) as demonstrated by agarose gel electrophoresis with pBR 322 DNA, ESR and laser flash photolysis measurements.

UV irradiation is the most frequent cause of DNA abnormality produced by reactive oxygen species (ROS).^{1,2} Photochemistry of aromatic amino acids has widely been studied with regard to generation of ROS such as singlet oxygen (¹O₂) and superoxide (O₂^{•-}).³⁻⁶ In general, there are two main processes responsible for photosensitized reactions (Scheme 1): (1) the chromophore is excited by light to a triplet state, and undergoes a direct electron or hydrogen exchange with a substrate, creating a free radical (Type I process); (2) energy transfer from the excited chromophore directly to oxygen, resulting in generation of ¹O₂ (Type II process).⁷⁻¹¹ Formation of ¹O₂ by UV irradiation of aromatic amino acids such as tryptophan and tyrosine, which absorb the light in the UV-B range (290–320 nm)¹² has been detected by its near IR emission at 1270 nm.^{13,14} The ¹O₂ quantum yields for tyrosine ($\Phi_{\Delta} = 0.14$) and tryptophan ($\Phi_{\Delta} = 0.062$) are significantly smaller than those of the corresponding triplet formation ($\Phi_{\text{T}} = 0.50$ and 0.18), respectively.¹⁴ Electron transfer from the triplet excited state to O₂ may contribute to the difference between the triplet quantum yield (Φ_{T}) and the ¹O₂ quantum yield (Φ_{Δ}).^{14,15} One of important

effects of ROS is abnormality *via* the DNA cleavage by ROS in the biological cell. However, DNA damage by ROS generated by photoirradiation of aromatic amino acid or neurotransmitter with oxygen has yet to be reported.

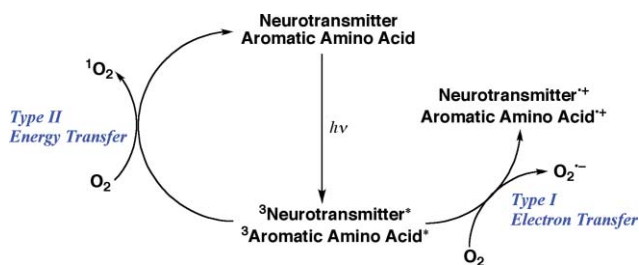
We report herein the effects of ROS produced by UV-B photoirradiation of a neurotransmitter, serotonin (5-hydroxytryptamine: 5-HT) and aromatic amino acids, tryptophan (Trp) and tyrosine (Tyr), with oxygen on DNA cleavage. The efficiency for the DNA cleavage with ROS produced by photoirradiation of a neurotransmitter and aromatic amino acids in Chart 1 was examined using agarose gel electrophoresis. We have also confirmed formation of superoxide ion by photoirradiation of 5-HT, which was most effective for DNA cleavage, by using electron spin resonance (ESR) and laser flash photolysis.



Chart 1 Neurotransmitter and aromatic amino acids used in this study.

The widely used assay with pBR322 supercoiled DNA^{15,16} was employed in order to clarify the active species responsible for the DNA cleavage activity upon UV-B irradiation of 5-HT, Trp and Tyr with O₂. The supercoiled DNA (form I) is efficiently cleaved into form II (nicked DNA) by photoirradiation of an air-saturated phosphate buffer solution (0.10 mol dm⁻³, pH 7.0) of 5-HT, Trp and Tyr with use of a monochromatized light ($\lambda = 300$ nm) from a xenon lamp as shown in Fig. 1 (lanes 5, 8 and 10, respectively). Serotonin exhibits the largest reactivity for the photoinduced DNA cleavage with oxygen. Without oxygen no photoinduced DNA cleavage was observed with 5-HT as shown in Fig. 1 (lane 4). Under dark conditions, no DNA cleavage occurs in the presence of 5-HT, Trp or Tyr as shown in Fig. 1 (lanes 3, 7 and 9, respectively).

To test the possible role of ¹O₂, the effect of ¹O₂ stabilizer was tested. When H₂O is replaced by D₂O, which is known to prolong the lifetime of ¹O₂,¹⁷ the DNA cleavage activity remains the same (ESI,† S1). This indicates that ¹O₂ does not play a significant role in the DNA-cleaving activity under UV-B irradiation. In contrast to the effect of the ¹O₂ stabilizer, the DNA cleaving activity was clearly inhibited by the addition of superoxide dismutase (SOD),¹⁸ which quenches O₂^{•-} (lane 6 in Fig. 1). This suggests that formation of O₂^{•-} plays an essential role in the DNA cleavage, although O₂^{•-} is generally regarded as a rather unreactive radical species.¹⁹



Scheme 1 Two possible pathways of generation of active species in the neurotransmitter and amino acid–O₂ system under UV-irradiation.

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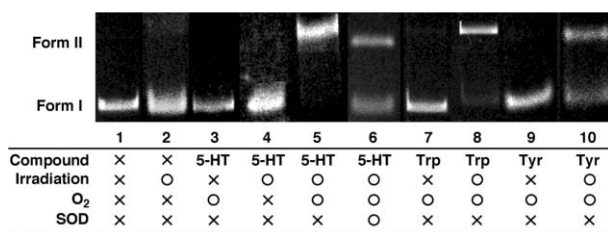


Fig. 1 DNA cleavage after 20 min photoirradiation with monochromatized light ($\lambda = 300$ nm). Lane 1, control; lane 2, photoirradiation; lane 3, no photoirradiation with 5-HT (1.0 mmol dm^{-3}); lane 4, photoirradiation of an Ar-saturated aqueous solution with 5-HT (1.0 mmol dm^{-3}); lane 5, photoirradiation of an O₂-saturated aqueous solution with 5-HT (1.0 mmol dm^{-3}); lane 6, photoirradiation of an O₂-saturated aqueous solution with 5-HT (1.0 mmol dm^{-3}) and SOD ($140 \mu\text{g dm}^{-3}$); lane 7, no photoirradiation of an O₂-saturated aqueous solution with Trp (1.0 mmol dm^{-3}); lane 8, photoirradiation of an O₂-saturated aqueous solution with Trp (1.0 mmol dm^{-3}); lane 9, no photoirradiation of an O₂-saturated aqueous solution with Tyr (1.0 mmol dm^{-3}); lane 10, photoirradiation of an O₂-saturated aqueous solution with Tyr (1.0 mmol dm^{-3}).

DNA can also be cleaved with 5-HT and Trp under irradiation of the UV-A light ($\lambda = 340$ nm) as shown in Fig. 2, where Tyr exhibits no reactivity because Tyr has no absorption at 340 nm.

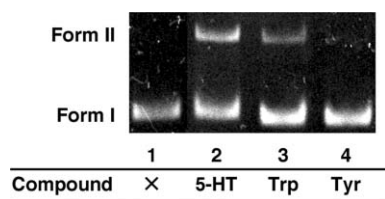


Fig. 2 Agarose gel electrophoresis of cleaved supercoiled pBR322 DNA after UVA-irradiation of aromatic amino acid and neurotransmitter with O₂ in an air-saturated aqueous buffer solution (pH 7.0) at 298 K. Cleavage of DNA after 20 min photoirradiation with monochromatized light ($\lambda = 340$ nm). Lane 1, control (DNA after photoirradiation); lane 2, photoirradiation with 5-HT (4.0 mmol dm^{-3}); lane 3, photoirradiation with Trp (4.2 mmol dm^{-3}); lane 4, photoirradiation with Tyr (4.5 mmol dm^{-3}).

Nanosecond laser flash photolysis measurements were performed to examine the photoinduced electron-transfer dynamics of 5-HT and aromatic amino acids with O₂. Transient absorption spectra of a phosphate buffer solution (0.10 mol dm^{-3} , pH 7.0) containing 5-HT ($1.7 \times 10^{-4} \text{ mol dm}^{-3}$) were observed upon the laser excitation at 266 nm (Fig. 3a). The shoulder absorption band at 480 nm is assigned to the triplet excited state of 5-HT (³5-HT*^{*}; * denotes the excited state) observed at 0.8 μs after laser excitation.²⁰ The broad absorption around 700 nm is attributed to hydrated electron (e_{aq}^-),¹⁷ which is produced by photoionization of 5-HT. The absorption band observed at 410 nm at 90 μs after laser excitation is assigned to the neutral 5-indoxyl radical that is formed by deprotonation of 5-HT⁺ due to the low $\text{p}K_{\text{a}}$ value of -0.7 .¹⁷ The decay time profile at 480 nm due to ³5-HT* obeyed first-order kinetics as shown in Fig. 3b.

In the presence of O₂, e_{aq}^- reacts immediately with O₂ by electron transfer to produce O₂^{•-}. On the other hand, the first-order decay rate constant of ³5-HT* increases with increasing concentration of O₂. The bimolecular quenching rate constant of ³5-HT* with O₂ is determined to be $9.2 \times 10^8 \text{ mol}^{-1} \text{ dm}^3 \text{ s}^{-1}$. This indicates that ³5-HT* is efficiently quenched by energy transfer or/and electron

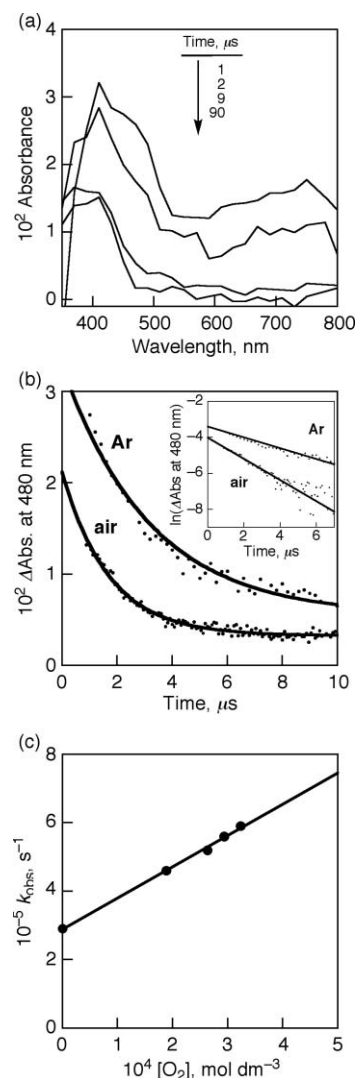


Fig. 3 (a) Transient absorption spectra of 5-HT ($2.1 \times 10^{-4} \text{ mol dm}^{-3}$) in a deaerated phosphate buffer solution (0.10 mol dm^{-3} , pH 7.0) after laser excitation at 266 nm. (b) Decay time profiles of ³5-HT* ($\lambda = 480$ nm) in argon- and air-saturated buffer solutions. The solid curves represent the best fit to the exponential decay. Inset: First order plots. (c) Plot of k_{obs} vs. $[\text{O}_2]$.

transfer from ³5-HT* to O₂. Formation of singlet oxygen by energy transfer from ³5-HT* to O₂ was previously confirmed by near-IR emission spectral measurements at 1270 nm from singlet oxygen.²⁰

The quenching rate constants (k_{q}) of the triplet excited states of Trp and Tyr were also determined by the decay of the T-T absorption, which is accelerated by the presence of oxygen (ESI, † S2, S3). The k_{q} values are listed in Table 1 together with the one-electron oxidation potentials of the ground state (E_{ox}) and the triplet excited states (³ E_{ox}^*) of Trp, 5-HT and Tyr. The E_{ox} values of ³Trp*, ³5-HT* and ³Tyr* are determined to be -1.84 , -1.94 and 2.25 V vs. SCE (saturated-calomel-electrode) by subtracting the triplet excitation energies (2.86, 2.82 and 3.18 eV)¹⁶ from the E_{ox} values of the ground states (1.02, 0.88 and 0.93 V vs. SCE).¹⁸ The k_{q} value increases with decreasing the E_{ox} value of the triplet excited state. This suggests that electron transfer from ³Trp*, ³5-HT* and ³Tyr* to O₂ may also play a role in the triplet quenching

Table 1 One-electron oxidation potentials at the ground and triplet excited states of neurotransmitter and amino acids (E_{ox} and ${}^3E_{\text{ox}}^*$, respectively), and the rate constants of electron transfer (k_{et}) from the triplet excited states to O_2

Compound	E_{ox} vs. SCE ^a /V	${}^3E_{\text{ox}}^*$ vs. SCE ^b /V	k_{et} ^c /mol ⁻¹ dm ³ s ⁻¹
Trp	1.02	-1.84	8.6×10^8
5-HT	0.88	-1.94	9.2×10^8
Tyr	0.93	-2.25	2.7×10^9

^a Taken from ref. 17. ^b Determined from phosphorescence reported in ref. 18, 23 and 24. ^c Experimental error is within 5%.

with O_2 . This is supported by the large driving force of electron transfer from ${}^3\text{Trp}^*$ (${}^3E_{\text{ox}}^* = -1.84$ V vs. SCE), ${}^35\text{-HT}^*$ (${}^3E_{\text{ox}}^* = -1.94$ V vs. SCE) and ${}^3\text{Tyr}^*$ (${}^3E_{\text{ox}}^* = -2.25$ V vs. SCE) to O_2 ($E_{\text{red}} = -0.40$ V vs. SCE).²¹ Thus, photoexcitation of 5-HT, Trp and Tyr with O_2 results in formation of $\text{O}_2^{\cdot-}$ via photoionization followed by electron transfer from e_{aq}^- to O_2 and also electron transfer from the triplet excited states to O_2 .²² ${}^3\text{Tyr}^*$ which has the most negative ${}^3E_{\text{ox}}^*$ value exhibits the highest reactivity toward O_2 (Table 1). However, no effective DNA cleavage was observed under UVA irradiation ($\lambda = 340$ nm) in Fig. 2, because Tyr has no absorption band at 340 nm. The formation of $\text{O}_2^{\cdot-}$ upon photoexcitation of 5-HT with O_2 by use of a mercury lamp is confirmed by the ESR spectrum measured at 77 K as shown in Fig. 4, where the anisotropic signals at $g_{\parallel} = 2.080$ and $g_{\perp} = 2.002$ are well assigned to $\text{O}_2^{\cdot-}$.^{25,26} 5-HT^+ is also observed around $g = 2.0042$,²⁷ which is overlapped with the g_{\perp} signal of $\text{O}_2^{\cdot-}$.

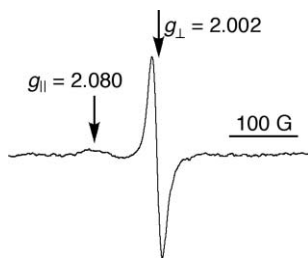


Fig. 4 ESR spectrum of $\text{O}_2^{\cdot-}$ generated by photoirradiation of an O_2 -saturated aqueous buffer solution (pH 7.0) containing 5-HT (0.47 mol dm^{-3}) with a high-pressure mercury lamp at 298 K and immediately measured at 77 K.

In summary, the photoreactivity of a neurotransmitter (serotonin) for DNA cleavage is significantly larger than aromatic amino acids (tryptophan and tyrosine) at ambient temperature. The photocleavage of DNA is initiated by electron transfer from the triplet excited states of a neurotransmitter and aromatic amino

acids to O_2 (Type I) rather than energy transfer to produce ${}^1\text{O}_2$ (Type II).

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